# **Pilot Scale Extraction and Fractionation of Rice Bran Oil Using Supercritical Carbon Dioxide**

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In the first stage of this two-stage process, crude rice bran oil was extracted with supercritical carbon dioxide (SC-CO<sub>2</sub>) from a 300 g batch of rice bran. Oil-laden SC-CO<sub>2</sub> from the extractor (24.1 MPa/ 40 °C) passed continuously to a second-stage column where an oil phase (raffinate) separated from the SC-CO<sub>2</sub> at various controlled temperatures and pressures. Measurement of the compositions of raffinates and extracts allowed calculation of partition coefficients of triglycerides, free fatty acids (FFAs),  $\alpha$ -tocopherol, sterols, and oryzanol and, hence, the selectivities of the fractionations. Fractionation removed almost all water and reduced the FFA concentration in raffinate by up to 50%. Oryzanol and  $\alpha$ -tocopherol concentrations in the raffinate were not reduced by fractionation, but the sterol concentration was reduced under conditions favoring FFA removal. Under the flow rate conditions studied (3.5 kg of CO<sub>2</sub>/h), the fractionations could be described by equilibria between oil and CO<sub>2</sub> phases.

Keywords: Supercritical carbon dioxide; extraction; fractionation; rice bran oil

### INTRODUCTION

Rice bran oil is a valuable domestic oil resource in many countries, although its acceptability as an edible oil has been hampered because of its commonly high free fatty acid (FFA) content resulting from the high lipase activity of the bran (Nicolosi et al., 1994). Deacidification by conventional means (alkali or physical refining) is a relatively expensive process and may not be economically feasible if alternative vegetable oils are readily available. Furthermore, the use of conventional refining methods can result in significant losses of minor but nutritionally important components of rice bran oil, such as oryzanol (Nicolosi et al., 1994). In a previous study, Shen et al. (1996) demonstrated the feasibility of extraction of oil from rice bran using supercritical CO<sub>2</sub> (SC-CO<sub>2</sub>) at various temperatures and pressures on single-stage pilot equipment and described some apparent partition coefficients for triglycerides, FFA,  $\alpha$ -tocopherol, sterols, and oryzanol in SC-CO<sub>2</sub> under the conditions studied. These results indicated that there may be potential to develop a fractionating extraction system through exploitation of these apparent SC-CO<sub>2</sub> solubility differences among various components.

Partial deacidification of some other vegetable oils by SC-CO<sub>2</sub> has been reported previously, for example: soybean oil (Friedrich et al., 1982), palm oil (Brunner and Peter, 1982), olive oils (Brunetti et al., 1989; Bondioli et al., 1992), and peanut oil (Ziegler and Liaw, 1993). After comparing the solubility isotherms of fatty acids and vegetable oil, Maheshwari et al. (1992) suggested that separation of fatty acids from triglycerides might be possible by using SC-CO<sub>2</sub> at densities <0.7 g/mL. Chrastil (1982) measured the solubility of certain fatty acids and triglycerides in SC-CO<sub>2</sub> within the pressure range 8-25 MPa and the temperature range 40-80 °C and showed that at certain temperatures and pressures, CO<sub>2</sub> has a higher solvent power for fatty acids than for the corresponding triglycerides. Zhao et al.

(1987) performed the fractional extraction of oil from rice bran by SC-CO<sub>2</sub> with a single column holding 20 g of rice bran, producing rice bran oil low in FFA using pressures of 15-35 MPa at 40 °C. However, the apparent solubilities of rice bran oil and the partition coefficients of its components, especially FFA, in SC- $CO_2$  of lower density (0.25–0.69 g/mL) have not been investigated. In a previous study (Shen et al., 1996), we explored the time course of extraction of rice bran oil and its components using dense CO<sub>2</sub> at various temperatures and pressures. While the partition coefficients measured in that study are of general utility in the design of oil extraction and refining procedures, the time course data are directly applicable only to the design of batch separations. The present study is an extension of that research and was conducted to determine the feasibility of using SC-CO<sub>2</sub> to simultaneously extract and fractionate oil from rice bran, with particular reference to deacidification, by means of a secondstage on-line solvent density reduction step.

More specifically, the aim of the present study was to continuously produce a rice bran oil of enhanced composition using a second-stage expansion column after primary SC-CO<sub>2</sub> extraction and to utilize the data from the expansion column to calculate the solubility of rice bran oil, the partition coefficients, and the selectivities of its components, as functions of temperature, pressure, and density under these lower density conditions.

#### MATERIALS AND METHODS

**Materials.** Medium-grain rice bran was provided by the Ricegrowers' Co-operative Ltd., Leeton, Australia. The moisture content and total hexane extractable matter in the bran were 8.48% and 17.59%, respectively.

**Extraction Methods.** A schematic diagram of the pilot plant extraction and fractionation unit (Distillers MG Ltd., U.K.) is shown in Figure 1. Food grade pure liquid carbon dioxide (CO<sub>2</sub>; 99.8% purity; CIG, Melbourne) was cooled and pressurized by a piston pump to a pressure of 24.1 MPa, which was regulated and checked by a variable pressure indicator controller. The pressurized CO<sub>2</sub> passed through a heater and

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**Figure 1.** Schematic diagram of the pilot scale  $CO_2$  extraction and fractionation plant used in this study, showing variable pressure indicator controllers (PIC), heaters (H), coolers (C), piston pumps (P), separation vessel (S), tailing column (TC), valves (V), and control valves (CV).

 Table 1. Distribution of Major Components of Rice Bran Oil, Extracted at 24.1 MPa/40 °C and Fractioned at Various

 Temperatures and Pressures<sup>a</sup>

used CO <sub>2</sub> (kg)	pressure (MPa)	temp (°C)	CO <sub>2</sub> density (g/mL)	amt of raffinate (g)	amt of extract (g)	raffinate FFA % <sup>b</sup>	extract FFA % <sup>b</sup>	concn of water in raffinate (%)	amt of water extracted (g)
13.1	8.6	50	0.2477	42.56	1.51	9.21	18.15	0.10	9.72
13.8	8.6	45	0.2804	41.37	2.03	7.39	68.9	0.08	10.02
14.0	8.6	40	0.3668	39.67	4.51	6.10	50.44	0.13	10.72
14.6	9.9	40	0.6242	35.31	9.13	5.34	33.61	0.14	10.34
13.9	11.2	40	0.6941	31.81	12.15	4.78	27.96	0.11	9.40

 $^a$  FFA % in hexane extractable oil is 9.79%.  $^b$  As oleic acid.

flowed up through a vertically mounted 1 L extractor equipped with a water jacket to maintain the set temperature of 40 °C. The extractor was loaded with 300.0 g of rice bran in each case, and each end was plugged with stainless steel mesh. The oil-laden CO<sub>2</sub> at 24.1 MPa/40 °C from the extractor passed through the valve into the fractionator, in which the pressure and temperature of the oil-laden CO<sub>2</sub> were held at the desired values by a back pressure regulator and a water jacket in which water was circulating through a water bath heater. On pressure reduction the CO<sub>2</sub> stream separated into an oil-rich phase and a CO2-rich phase, with the oil-rich phase precipitating to the bottom of the fractionator. The CO<sub>2</sub>-rich phase from the fractionation column flowed through a separation vessel with a glass window, where SC-CO2 was depressurized and vented through a packed tailing column and vaporizer, leaving the extract in the separation vessel. The raffinate was collected from the bottom of the fractionator. The CO<sub>2</sub> flow rate was manually adjusted to an average of 3.5 kg/h. Under the above conditions, extraction and fractionation were continued for 4 h simultaneously.

**Analytical Methods.** The amounts of total extract and raffinate were determined gravimetrically. Moisture in the extract and raffinate was measured by the vacuum oven method (AOAC, 1990). The methods of analysis for FFA, oryzanol, tocopherols, and sterols were as described by Shen et al. (1996).

#### **RESULTS AND DISCUSSION**

**Isothermal Fractionation at 40** °C. The effect of the pressure in the fractionation column on the distribution of the major components of rice bran oil between the raffinate and extract is shown in Table 1. As expected, the increased  $CO_2$  density, resulting from increased pressure at constant temperature of 40 °C, increased the solvent power of the SC-CO<sub>2</sub> for all components except water and thus increased the mass

of the extract at the expense of the raffinate. In the case of water, because only a small fraction remained in the raffinate under all conditions studied, it was not possible to detect any trends in the distribution of water between oil and  $CO_2$  phases. The second-stage fractionation step left only about 0.1% water in the raffinate. This value is well below the specified maximum of 0.3% water for many vegetable oils of similar composition (AFSC, 1996) and is a very useful result because the removal of water from rice bran oil increases its stability and commercial value. This is a great improvement on the single-stage  $CO_2$  extraction at 24 MPa/40 °C reported by Shen et al. (1996), which produced rice bran oil containing about 20% water.

Bondioli et al. (1992) have refined lampante olive oil in a SC-CO<sub>2</sub> extraction plant operating in continuous countercurrent mode and reported the influence of pressure on FFA/triglyceride separation with pressures of 8.0, 9.0, and 11.0 MPa at 40 °C. They obtained decreased refined oil yield, decreased FFA concentration in refined oil, and a decreasing and then increasing trend for FFA concentration in the extract, as pressure was increased. Our fractionation trends are similar to those reported by Bondoli et al. (1992) except for FFA concentrations in the extracts (Table 1).

Zhao et al. (1987) fractionally extracted rice bran oil from 20 g of rice bran, which consumed 3.5 kg of  $CO_2$  at pressures in steps from 15–35 MPa/40 °C. Combining the fractions collected after the first pressure step produced an oil low in FFA. The oil low in FFA contained 84.5% of rice bran oil, 50.0% of FFA, 81.8% of oryzanol, and 84.5% of tocopherols in the total extract. This result can be compared with the present results at 11.2 MPa/40 °C of the fractionation condition, for

Table 2. Distribution of Minor Components of Rice Bran Oil, Extracted at 24.1 MPa/40 °C and Fractionated at Various Temperatures and Pressures

pressure temp		oryzanol (mg/g of oil)		campesterol (mg/g of oil)		stigmasterol (mg/g of oil)		eta-sitosterol (mg/g of oil)		α-tocopherol (mg/g of oil)	
(MPa)	(°C)	raffinate	extract	raffinate	extract	raffinate	extract	raffinate	extract	raffinate	extract
8.6	50	10.85	ne	1.80	ne	1.60	ne	1.40	ne	0.25	ne
8.6	45	12.06	5.20	1.75	1.49	1.55	1.29	9.89	7.10	0.24	nd
8.6	40	11.20	6.43	1.65	3.40	1.50	3.10	9.30	18.50	0.22	0.00
9.9	40	12.15	6.36	1.45	2.70	1.25	2.45	7.76	15.09	0.19	0.10
11.2	40	12.65	5.43	1.45	2.35	1.20	2.15	6.90	13.48	0.19	0.23

<sup>a</sup> ne, not enough sample to detect; nd, not detected.

which the respective recoveries were 72.3%, 31.0%, 85.9%, and 68.0%. The present method has achieved a greater reduction in FFA than that by the method of Zhao et al. (1987). Furthermore, the consumption of  $CO_2$  per gram of rice bran in the present method was only 26% of the consumption in the work of Zhao et al. (1987).

In the present study, after 13.9 kg of CO<sub>2</sub> had been used, only 83.39% of hexane extractable oil was recovered, whereas in our previous extraction trials (Shen et al., 1996) 92.97% of hexane extractable oil was recovered after 13.2 kg of CO<sub>2</sub> was consumed. Since, in the present study, a higher average flow rate of 3.5 kg/h of SC-CO<sub>2</sub> was used rather than the 2.5 kg/h of  $SC-CO_2$  in our former work (Shen et al., 1996), one possible explanation for these different results is that after about 75% of hexane extractable oil has been recovered, mass transfer difficulties become important and the yield of oil decreases with increased  $CO_2$  flow rate. In other words, at the higher  $CO_2$  flow rate the later stages of extraction deviate more from equilibrium since the contact time between the solvent and the oil is reduced.

The distribution of other oil components between the raffinate and extract is shown in Table 2. These mostly followed the previously reported trends in the partition coefficients between oil and SC-CO<sub>2</sub> phases (Shen et al., 1996). The sterols (campesterol, stigmasterol, and  $\beta$ -sitosterol) were concentrated in the extract. Oryzanol was preferentially distributed to the raffinate, which is consistent with the respective partition coefficients reported by Shen et al. (1996). However, it appears that in the present study, because of the lower densities of CO<sub>2</sub> at 8.6 and 9.9 MPa,  $\alpha$ -tocopherol was concentrated in the raffinate and not preferentially extracted as occurred with CO<sub>2</sub> at higher densities (Shen et al., 1996).

Isobaric Fractionation at 8.6 MPa. Table 1 shows the effect of temperature in the fractionation column on the distribution of the major components of rice bran oil between the raffinate and extract at 8.6 MPa. By increasing the fractionation temperature from 40 to 45 °C and then to 50 °C, the SC-CO<sub>2</sub> density and hence its solvent power were progressively decreased. This resulted in a progressively increasing yield of raffinate and corresponding decreasing yield of extract. The FFA concentration in the raffinate increased with increasing temperature, while the FFA concentration in extract initially increased and then decreased. In SC-CO<sub>2</sub> fractionation of olive oil, Bondioli et al. (1992) reported that with temperatures of 40 and 60 °C at a pressure of 13 MPa there was an increase in raffinate yield and FFA concentration in both the raffinate and extract at the higher temperature. The trends in our fractionation are in accord with this study except for the reduced FFA concentration in the rice bran oil extract at higher temperatures. Table 2 shows the effect of the temperature in the fractionation column on the distribution of rice bran oil minor components between the refined oil fraction and extract. When the temperature was increased from 40 to 45 °C and then to 50 °C at constant pressure, resulting in decreased density of  $CO_2$ , the concentration of all minor components increased in the refined oil with the exception of oryzanol. The reason for this behavior of oryzanol is still uncertain. The densities of  $CO_2$  at 50 °C/8.6 MPa were too low to produce sufficient material for analysis of any extract component other than FFA.

**Selectivities and Partition Coefficients.** In our previous paper on the extraction of rice bran oil in dense  $CO_2$  (Shen et al., 1996), the results were presented as partition coefficients calculated on a mass fraction basis. In the present study, ratios of the partition coefficients are used to derive "selectivity" (*S*), which provides a useful basis for comparison with respect to the fractionation process. *S* values were calculated for different extraction conditions according to the equation (Brunetti, 1989)

$$S = \frac{W_{\rm E}^{\rm I}/W_{\rm R}^{\rm I}}{W_{\rm F}^{\rm T}/W_{\rm R}^{\rm T}}$$

where  $W_{\rm E}$  and  $W_{\rm R}$  are the weight fraction of the component (I) in the extract and refined oil and  $W_{\rm E}^{\rm T}$ and  $W_{\rm R}^{\rm T}$  the weight fraction of triglycerides in the extract and refined oil, respectively. The "separation factors" calculated by Arul et al. (1994) are actually ratios of partition coefficients calculated on a mole fraction basis. Brunetti (1989) had earlier used the term "distribution coefficient or solvent selectivity" to refer to a ratio of partition coefficients calculated on a mass fraction basis. Nilsson et al. (1991, 1992) have consistently used partition coefficients calculated on a mass fraction basis and have defined the term "selectivity" as the ratio of such partition coefficients.

Table 3 shows the selectivities for some minor rice bran oil components isothermally at 40 °C and isobarically at 8.6 MPa, respectively. In every case, FFA had the highest selectivities, meaning that the FFA were preferentially enriched in the extract under all  $CO_2$ conditions used, compared with the other components. The selectivities of oryzanol decreased with increasing  $CO_2$  density, meaning that the best separation of oryzanol from triglycerides was obtained at 11.2 MPa/ 40 °C among all of the conditions used, while  $\alpha$ -tocopherol was preferentially retained in the refined oil under most conditions.

Figure 2 plots the partition coefficients versus  $CO_2$  density for triglycerides, FFA, sterols, and oryzanol. Data from a previous study (Shen et al., 1996) are included with data from the present work. The data from Shen et al. (1996) were determined at high pressure at which the assumed equilibrium condition is approached by passing  $CO_2$  over dispersed oil droplets in rice bran. In the present study data were determined

Table 3. Selectivities for Some Minor Components of CO<sub>2</sub> Extracted Rice Bran Oil

pressure (MPa)	temp (°C)	FFA	oryzanol	campesterol	stigmasterol	$\beta$ -sitosterol	α-tocopherol	
8.6	50	2.23	ne <sup>a</sup>	ne	ne	ne	ne	
8.6	45	27.82	1.29	2.55	2.49	2.14	< 0.06	
8.6	40	15.64	1.08	3.90	3.91	3.76	0.02	
9.9	40	9.00	0.75	2.66	2.80	2.78	0.75	
11.2	40	7.73	0.57	2.14	2.36	2.58	1.62	
<sup>a</sup> ne, not enough sample to detect.								

Table 4.	<b>Comparison</b>	of Partition	Coefficients	of Triglyceri	des with	Solubility	of Vegetable Oil

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CO <sub>2</sub> density (g/mL) temp (°C) pressure (MPa)	0.6941 40 11.2	0.7767 60 24.1	0.8087 40 17.2	0.8406 40 20.0	0.8732 40 24.1	0.9159 40 31.0
partition coefficient of triglycerides ( $\times 10^3$ ) solubility of vegetable oil and triolein (g/kg of CO <sub>2</sub> )	$0.66^{a}$ $0.6^{b}$	${3.52^a(3.1)^e}\over{3.1^e}$	$2.50^{a}$ $2.0^{b}$	$2.9^{c} (3.1)^{d}$	$5.52^{a}$ $5.0^{b}$	$rac{6.93^{a}}{7.2^{b}}$

<sup>a</sup> This work. <sup>b</sup> Fattori et al. (1988) (Figure 5 for canola oil). <sup>cd</sup> Stahl et al. (1980) (Figure 4 for sunflower oil and Figure 5 for soybean oil). <sup>e</sup> Nilsson et al. (1991) solubility and partition coefficient of pure triolein.



**Figure 2.** Partition coefficients for some components of rice bran oil at 40 °C plotted on a logarithmic scale. Single points represent the mean values of data from duplicated experiments.

at lower pressures, at which the assumed equilibrium is approached by condensing oil from solution in  $CO_2$ . The two sets of data are approximately contiguous, indicating that the assumption of equilibrium is justified in both sets of experiments.

The selectivities are ratios of partition coefficients and determine the maximum possible degree of separation of any pair of components under given conditions in a simple batch or cocurrent process. For optimal separation the selectivity must be maximized (or minimized, depending on how it is defined). This condition is best shown by maximum distance between the respective partition coefficient curves when plotted on a logarithmic scale. Figure 2 has been plotted on a logarithmic scale to illustrate the possibilities and limitations of  $CO_2$  as a fractionating solvent for rice bran oil.

The vertical distances between the curves clearly show that FFA are best separated from triglycerides at low CO<sub>2</sub> densities (<0.7 g/mL), which is in agreement with the data of Maheshwari et al. (1992). The curves for sterols are all very close to each other, indicating the poor ability of CO<sub>2</sub> alone to separate the sterols from each other at 40 °C in the CO<sub>2</sub> density range tested. This could be expected because of the similarities of the molecular structures of these sterols. Since the sterol curves are located between the FFA and triglyceride curves at  $CO_2$  densities <0.8 g/mL, it is inevitable that any simple process to separate FFA from triglycerides at 40 °C and a CO<sub>2</sub> density of <0.8 g/mLwill also partially remove sterols from the triglycerides. In other words, at 40 °C, to preserve the sterol content of rice bran oil, it is necessary to conduct the deacidification at densities higher than optimal. It is possible that



**Figure 3.** Partition coefficients of triglycerides in rice bran oil plotted on a logarithmic scale. Single points represent the mean values of data from duplicated experiments.

another temperature could be more favorable or that the use of entrainers or adsorbents could overcome this difficulty.

Oryzanol had a partition coefficient that was less than or equal to the partition coefficient of triglycerides under all  $CO_2$  conditions at 40 °C. At  $CO_2$  densities >0.6 g/mL it would be possible to separate rice bran oil into a high oryzanol fraction and a low oryzanol fraction. The high oryzanol fraction would also inevitably have a reduced FFA content compared with the unfractionated oil.

In the present study, partition coefficients were also measured at temperatures of 45 and 50 °C and a CO<sub>2</sub> pressure of 8.6 MPa. These conditions produced extremely low partition coefficients that are outside the useful range. However, our earlier work (Shen et al., 1996) included some measurements at 60, 20, and 0 °C using higher CO<sub>2</sub> densities, which show the direction and magnitude of temperature effects. These measurements are included with the triglyceride and FFA partition coefficient isotherms of the present study in Figures 3 and 4, respectively. The use of density rather than pressure as the independent variable simplifies the understanding of the temperature effect. This approach has also been taken by del Valle and Aguilera (1988), who measured and compiled the solubility data of vegetable oil in SC-CO<sub>2</sub> over a range of temperatures from 20 to 80 °C and plotted them on a logarithmic scale against  $CO_2$  density to show a family of parallel straight lines. The solubility isotherms of canola oil in CO<sub>2</sub> as a function of CO<sub>2</sub> density at temperatures from 25 to 70 °C measured by Fattori et al. (1988) displayed four parallel curves. Maheshwari et al. (1992) reported the predicted solubilities of five free fatty acids as a function of temperature and density of SC-CO<sub>2</sub>. When plotted



**Figure 4.** Partition coefficients of FFA in rice bran oil plotted on a logarithmic scale. Single points represent the mean values of data from duplicated experiments.

on a logarithmic scale, these appeared as a group of parallel straight lines in all cases. The data plotted in Figures 3 and 4 of the present study are consistent with the partition coefficient isotherms being families of parallel curves. For comparison, Fattori et al. (1988) also plotted their data using pressure as the independent variable and showed that the isotherms exhibited crossover points. Yun et al. (1991) reported that solubility curves of cholesterol in CO2 plotted on a logarithmic scale versus CO2 density showed a parallel linear trend of data at temperatures from 40 to 60 °C. Examination of the data points of Figure 3 of the present study indicates a complex relationship between isotherms if they were plotted against pressure. The use of density as the independent variable is therefore recommended for studies of isotherms in supercritical fluids.

Partition Coefficients and Solubilities. Any data on total solubility of a relatively homogeneous mixture carry the assumption that the measured solute is present in the oil phase at 1000 g/kg. Thus, if solubility data are expressed in grams per kilogram of CO<sub>2</sub>, they can be converted to partition coefficients for comparison with the present results by dividing by 1000. A similar treatment of the connection between measured solubilities and partition coefficients was described by Bamberger et al. (1988), who used partition coefficients based on mole fractions. Table 4 compares the present results with those from the literature on vegetable oil solubilities. The partition coefficients for triglycerides measured in this study and our previous study (Shen et al., 1996) can thus be directly related to the extensive literature on the solubility of vegetable oils (mixtures of triglycerides) in dense  $CO_2$  (Brunetti et al., 1989; Eggers et al., 1985; Lee et al., 1986; Stahl et al., 1980).

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## LITERATURE CITED

AFSC. Standard G1: Edible fats and oils. In *Australian Food Standards Code*; Australian Government Publishing Service: Canberra, Nov 1996.

- AOAC. Method 926.12: Moisture and volatile matter in oils and fats. In *Official Methods of Analysis of the Association of Official Analytical Chemists*, 15th ed.; Helrich, K., Ed.; AOAC: Arlington, VA, 1990.
- Arul, J.; Tardif, R.; Boudreau, A.; McGinnis, D. S.; Lencki, R.
   W. Solubility of milk fat triglycerides in supercritical carbon dioxide. *Food Res. Int.* 1994, *27*, 459–467.
- Bamberger, T.; Erickson, J. C.; Cooney, C. L. Measurement and model prediction of solubilities of pure fatty acids, pure triglycerides, and mixtures of triglycerides in supercritical carbon dioxide. *J. Chem. Eng. Data* **1988**, *33*, 327–333.
- Bondioli, P.; Mariani, C.; Lanzani, A.; Fedeli, E.; Mossa, A.; Muller, A. Lampante olive oil refining with supercritical carbon dioxide. J. Am. Oil Chem. Soc. 1992, 69, 477–480.
- Brunetti, L.; Daghetta, A.; Fedeli, E.; Kikic, I.; Zanderighi, L. Deacidification of olive oils by supercritical carbon dioxide. *J. Am. Oil Chem. Soc.* **1989**, *66*, 209–217.
- Brunner, G.; Peter, S. On the solubility of glycerides and fatty acids in compressed gases in the presence of an entrainer. *Sep. Sci. Technol.* **1982**, *17*, 199–214.
- Chrastil, J. Solubility of solids and liquids in supercritical gases. J. Phys. Chem. **1982**, 86, 3016-3021.
- del Valle, J. M.; Aguilera, J. M. An improved equation for predicting the solubility of vegetable oils in supercritical CO<sub>2</sub>. Ind. Eng. Chem. Res. **1988**, 27, 1551–1553.
- Eggers, R.; Sievers, U.; Stein, W. High pressure extraction of oil seed. J. Am. Oil Chem. Soc. **1985**, 62, 1222–1230.
- Fattori, M.; Bulley, N. R.; Meisen, A. CO<sub>2</sub> extraction of canola seed: oil solubility and effect of seed treatment. *J. Am. Oil Chem. Soc.* **1988**, *65*, 968–974.
- Friedrich, J. P.; List, G. R. Characterisation of soybean oil extracted by supercritical carbon dioxide and hexane. *J. Agric. Food Chem.* **1982**, *30*, 192–193.
- Lee, A. K. K.; Bulley, N. R.; Fattori, M.; Meisen, A. Modelling of supercritical carbon dioxide extraction of canola oilseed in fixed beds. J. Am. Oil Chem. Soc. 1986, 63, 921–925.
- Maheshwari, P.; Nikolov, Z. L.; White, T. W.; Hartel, R. Solubility of fatty acids in supercritical carbon dioxide. *J. Am. Oil Chem. Soc.* **1992**, *69*, 1069–1076.
- Nicolosi, R. J.; Rogers, E. J.; Ausman, L. M.; Orthoefer, F. T. Rice bran oil and its health benefits. In *Rice Science and Technology*; Marshall, W. E., Wadsworth, J. I., Eds.; Dekker: New York, 1994.
- Nilsson, W. B.; Gauglitz, E. J.; Hudson, J. K. Solubilities of methyl oleate, oleic acid, oleyl glycerols, and oleyl glycerol mixtures in supercritical carbon dioxide. *J. Am. Oil Chem. Soc.* **1991**, *68*, 87–91.
- Nilsson, W. B.; Seaborn, G. T.; Hudson, J. K. Partition coefficients for fatty acid esters in supercritical fluid  $CO_2$  with and without ethanol. *J. Am. Oil Chem. Soc.* **1992**, *69*, 305–308.
- Shen, Z.; Palmer, M. V.; Ting, S. S. T.; Fairclough, R. J. Pilot scale extraction of rice bran oil with dense carbon dioxide. *J. Agric. Food Chem.* **1996**, *44*, 3033–3039.
- Stahl, E.; Schutz, E.; Mangold, K. Extraction of seed oils with liquid and supercritical carbon dioxide. J. Agric. Food Chem. 1980, 28, 1153–1157.
- Yun, J. S. L.; Liong, K. K.; Gurdial, G. S.; Foster N. R. Solubility of cholesterol in supercritical carbon dioxide. *Ind. Eng. Chem. Res.* **1991**, *30*, 2476–2482.
- Zhao, W.; Shishikura, A.; Fujimoto, K.; Arai, K.; Saito, S. Fractional extraction of rice bran oil with supercritical carbon dioxide. *Agric. Biol. Chem.* **1987**, *51*, 1773–1777.
- Ziegler, G. R.; Liaw, Y. J. Deodorization and deacidification of edible oils with dense carbon dioxide. J. Am. Oil Chem. Soc. 1993, 70, 947–953.

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